

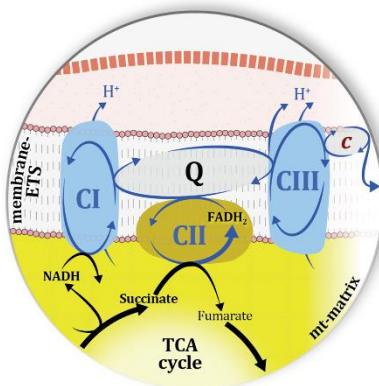
Theoretical Communication

Cite

Gnaiger E (2023) Complex II ambiguities – FADH₂ in the electron transfer system. MitoFit Preprints 2023.3.
<https://doi.org/10.26124/mitofit:2023-0003>

Conflicts of interest

The author declares no conflict of interest.



Received 2023-03-24

Accepted 2023-03-24

Online 2023-03-24

Keywords

coenzyme Q junction, Q-junction
 Complex II, CII
 electron transfer system, ETS
 fatty acid oxidation, FAO
 flavin adenine dinucleotide,
 FADH₂/FAD
 succinate dehydrogenase, SDH
 tricarboxylic acid cycle, TCA

Complex II ambiguities – FADH₂ in the electron transfer system

 Erich Gnaiger

Oroboros Instruments, Innsbruck, Austria.

Correspondence: erich.gnaiger@oroboros.at

Summary

The current narrative that the reduced coenzymes NADH and FADH₂ feed electrons from the tricarboxylic acid (TCA) cycle into the mitochondrial electron transfer system can create ambiguities around respiratory Complex CII. Succinate dehydrogenase or CII reduces FAD to FADH₂ in the canonical forward TCA cycle. However, some graphical representations of the membrane-bound electron transfer system (ETS) depict CII as the site of oxidation of FADH₂. This leads to the false belief that FADH₂ generated by electron transferring flavoprotein (CTF) in fatty acid oxidation and mitochondrial glycerophosphate dehydrogenase (CGpDH) feeds electrons into the ETS through CII. In reality, NADH and succinate produced in the TCA cycle are the *substrates* of Complexes CI and CII, respectively, and the reduced flavin groups FMNH₂ and FADH₂ are downstream *products* of CI and CII, respectively, carrying electrons from CI and CII into the Q-junction. Similarly, CTF and CGpDH feed electrons into the Q-junction but not through CII. The ambiguities surrounding Complex II in the literature call for quality control, to secure scientific standards in current communications on bioenergetics and support adequate clinical applications.

The tricarboxylic acid (TCA) cycle – the citric acid cycle or Krebs cycle – sparked a renaissance of interest in cellular and mitochondrial bioenergetics (Gnaiger et al 2020; Arnold, Finley 2023). TCA cycle metabolites are (1) oxidized while reducing NAD⁺ to NADH in the forward cycle, or (2) transported into the cytosol (Murphy MP, O'Neill LAJ 2018). Succinate dehydrogenase (succinate:quinone oxidoreductase, Complex CII) plays a key role in metabolic remodeling in cancer tissue (DeBerardinis, Chandel 2016; Schöpf et al 2020). The reversed TCA cycle has gained interest in studies ranging from anaerobic metabolism (Hochachka, Somero 2002), thermodynamic efficiency of anoxic and aerobic ATP production (Gnaiger 1993), reversed electron transfer and ROS production (Tretter et al 2016; Robb et al 2018), hypoxia and ischemia-reperfusion injury (Couchani et al 2014), to pathway evolution (Lane 2022).

Complex CII participates both in the membrane-bound electron transfer system (membrane-ETS) and TCA cycle (matrix-ETS plus CII; Gnaiger et al 2020). Branches of electron transfer from the reduced coenzyme NADH of nicotinamide adenine dinucleotide N and succinate S converge at coenzyme Q (Q-junction; Figure 1a).

The reduced flavin groups FADH₂ of flavin adenine dinucleotide and FMNH₂ of flavin mononucleotide are at functionally comparable levels in the electron transfer to Q from CII and CI, respectively, just as succinate and NADH are the comparable reduced substrates of CII and CI, respectively (Hatefi 1962; Tzagoloff 1982; Gnaiger 2020). In CII the oxidized form FAD is reduced by succinate to the product FADH₂ while fumarate is formed as the oxidized product in the TCA cycle. In CI FMN is reduced by NADH forming (red) FMNH₂ and (ox) NAD⁺. FADH₂ and FMNH₂ are reoxidized downstream in CII and CI by electron transfer to Q in the membrane-bound ETS (Figure 1b).

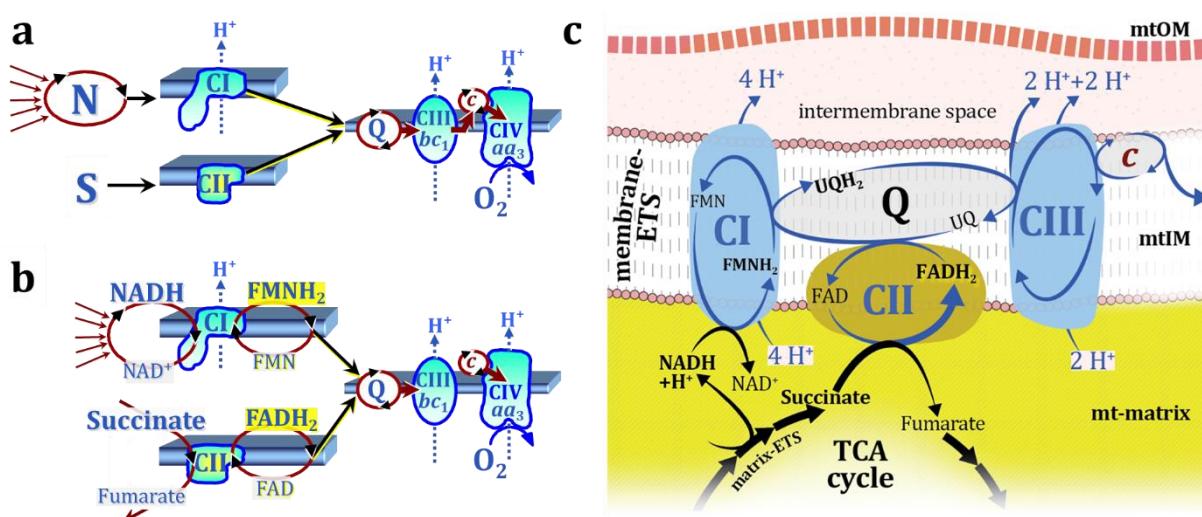


Figure 1. Convergent electron transfer from NADH and succinate to the Q-junction. (a) Electron flow catalyzed by dehydrogenases localized in the mitochondrial (mt) matrix reduces nicotinamide adenine dinucleotide N and converges at the N-junction. Electron flow through Complexes CI and CII converges at the Q-junction. Modified from Gnaiger (2020). (b) NADH and succinate are substrates of CI- and CII-catalyzed redox reactions, respectively. FMNH₂ and FADH₂ are products in CI and CII, respectively. (c) Complex CII is integrated in the membrane-bound electron transfer system (membrane-ETS in the mt-inner membrane mtIM) and the TCA cycle (matrix-ETS). Electron flow [2 H] + 0.5 O₂ → H₂O from succinate reduces FAD to FADH₂. [2 H] from FADH₂ reduces ubiquinone UQ to ubiquinol UQH₂. Complex CIII passes electrons to cytochrome c.

1. The source and consequence of Complex II ambiguities

'No representation is ever perfectly expressive, for if it were it would not be a representation but the thing itself' (Grosholz 2007). Ambiguities emerge if the representation of a concept is vague to an extent that allows for equivocal interpretations. As a consequence, even a basically clear concept (Figure 1c) may be communicated as a divergence from an established truth. The following quotes from Cooper (2000) provide an example (Figure 2).

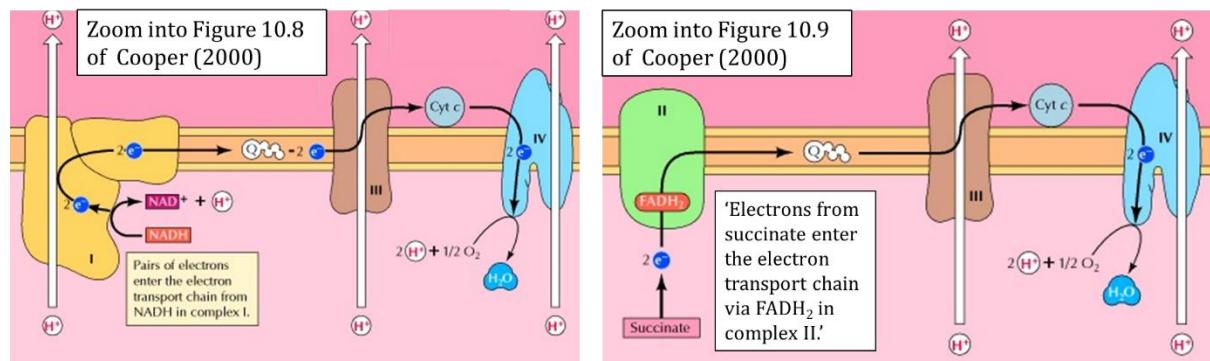


Figure 2. Electron flow into Complexes CI (left) and CII (right). Zoom into figures of Cooper (2000).

(1) The standard comparison is made between NADH (linked to CI) and FADH₂ (linked to CII): '*Electrons from NADH enter the electron transport chain in complex I, .. A distinct protein complex (complex II), which consists of four polypeptides, receives electrons from the citric acid cycle intermediate, succinate (Figure 10.9). These electrons are transferred to FADH₂, rather than to NADH, and then to coenzyme Q.*'

(2) '*In contrast to the transfer of electrons from NADH to coenzyme Q at complex I, the transfer of electrons from FADH₂ to coenzyme Q is not associated with a significant decrease in free energy and, therefore, is not coupled to ATP synthesis.*' Note that CI is in the path of electron transfer from NADH to coenzyme Q. In contrast, electron transfer from FADH₂ to coenzyme Q is downstream of CII. Thus even a large Gibbs force ('*decrease in free energy*') in FADH₂→Q would fail to drive the coupled process of proton translocation through CII. The Gibbs force in S→FADH₂ must be accounted for. (In parentheses: None of these steps are directly coupled to ATP synthesis. Redox-driven proton translocation must be distinguished from phosphorylation of ADP driven by the protonmotive force).

(3) CII receives electrons from succinate, yet it is suggested that '*electrons from succinate enter the electron transport chain via FADH₂ in complex II.*' The ambiguity is caused by a lack of unequivocal definition of the electron transfer system (*electron transport chain*). Two contrasting definitions are implied of the '*electron transport chain*' or ETS. (a) CII is part of the ETS. Hence electrons enter the ETS from succinate but not from FADH₂ – from the matrix-ETS to the membrane-ETS (Figure 1c). (b) If electrons enter the '*electron transport chain via FADH₂ in complex II*', then CII would be upstream and hence not part of the ETS (to which conclusion obviously nobody would agree). There remains the ambiguity of electron entry into CII from succinate (Figure 1) or from FADH₂ as the product of succinate dehydrogenase in the TCA cycle (Figure 3).

2. The FADH₂ - FAD confusion in the succinate-pathway

The narrative that the reduced coenzymes NADH and FADH₂ feed electrons from the TCA cycle into the mitochondrial electron transfer system causes confusion. As a consequence, FADH₂ appears in several publications erroneously as the substrate of CII in the ETS linked to succinate oxidation. This error is widely propagated (Supplement S1 and S2) and requires clarification (Gnaiger 2020; page 48). The following examples illustrate the transition from ambiguity to error.

(1) Ambiguities in graphical representations, where FADH₂ is the product and substrate of CII in the same figure (Figure 3).

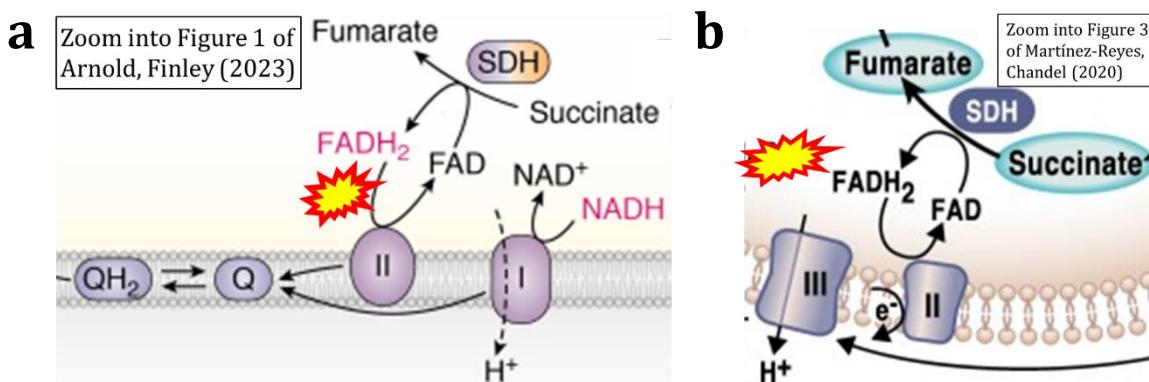
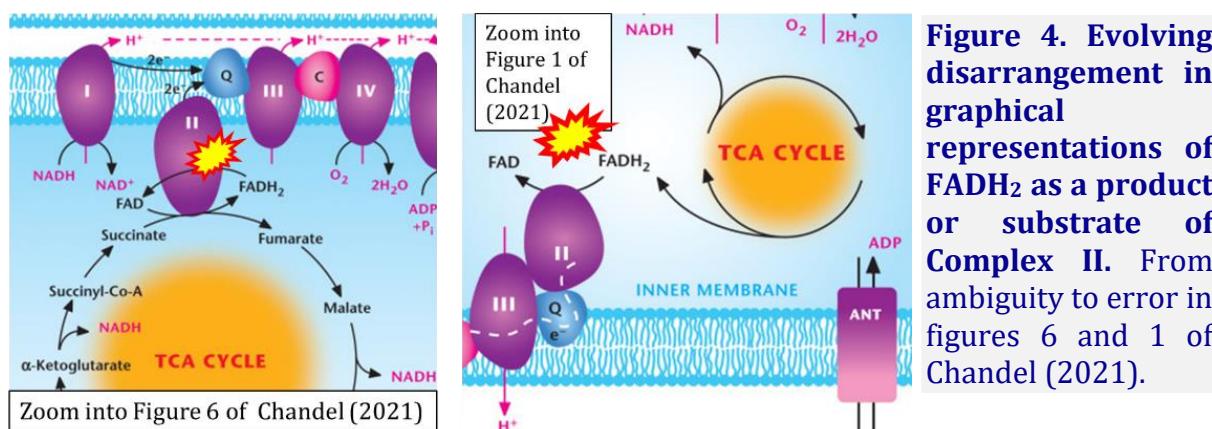


Figure 3. FADH₂ depicted as product and substrate of Complex II. Zoom into figures by (a) Arnold, Finley (2023) and (b) Martínez-Reyes, Chandel (2020).



(2) Evolution from ambiguity to error in graphical representations (Figure 4).

(3) Discrepancy between erroneous graphical representation (Figure 5) and correct text. 'Reducing equivalents (NADH, FADH₂) provide electrons that flow through complex I, the ubiquinone cycle (Q/QH₂), complex III, cytochrome c, complex IV, and to the final acceptor O₂ to form water' (Fisher-Wellman, Neufer 2012).

(4) Simple graphical errors (Figure 6).

(5) Error propagation from graphical representation (Figure 3a) to text: 'SDH reduces FAD to FADH₂, which donates its electrons to complex II'; 'each complete turn of the TCA cycle generates three NADH and one FADH₂ molecules, which donate their electrons to complex I and complex II, respectively'; 'complex I and complex II oxidize NADH and FADH₂, respectively' (Arnold, Finley 2023).

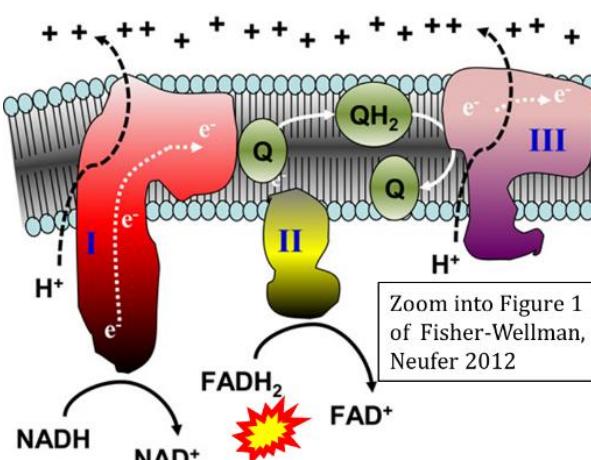


Figure 5. FADH₂ is shown as the substrate of Complex II. This graphical representation contradicts the text that clarifies that FADH₂ provides electron flow through the Q-cycle (Fisher-Wellman and Neufer 2012).

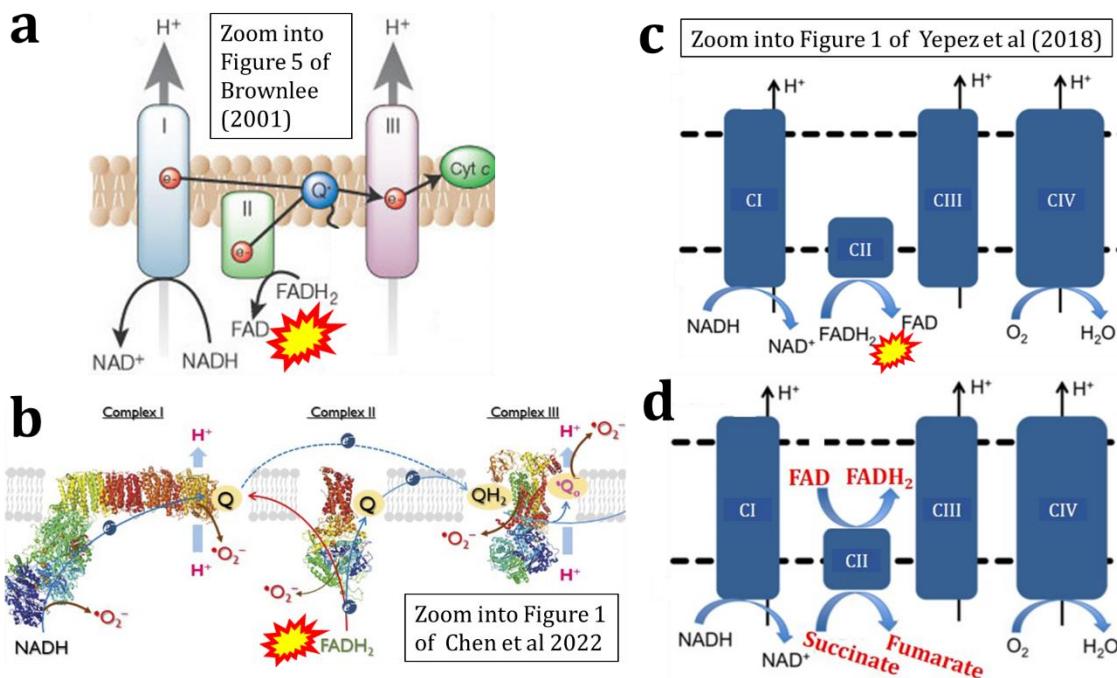


Figure 6. FADH₂ is shown as the substrate of Complex II. Zoom into figures from **(a)** Brownlee (2001), **(b)** Chen et al (2022), **(c)** Yépez et al (2018). **(d)** Correction showing succinate and FAD as substrates of Complex II.

The presentation of electron transfer from FADH₂ to CII (Figure 6; Supplement Figures S1 and S2) has a logical consequence. Electron transferring flavoprotein in β-oxidation and mitochondrial glycerophosphate dehydrogenase generate FADH₂. If FADH₂ would donate electrons to CII, then CII can be seen as an enzyme involved downstream of FADH₂ in FAO and the glycerophosphate shuttle. This topic requires clarification.

3. Complex II and fatty acid oxidation

Electron transferring flavoprotein CETF and CI are the respiratory Complexes involved in convergent electron entry into the Q-junction during FAO (Figure 7).

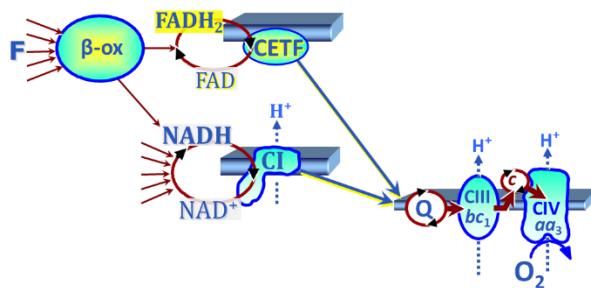


Figure 7. Fatty acid oxidation through the β-oxidation cycle (β-ox), electron transferring flavoprotein CETF, and Complex CI with convergent electron transfer into the Q-junction. Modified after Gnaiger (2020).

In the β-oxidation cycle of FAO, acetyl-CoA and the reducing equivalents FADH₂ and NADH are formed in reactions catalyzed by acyl-CoA dehydrogenases and hydroxyacyl-CoA dehydrogenases, respectively, in the mitochondrial matrix (Houten et al 2016). When FADH₂ is erroneously shown as a substrate of CII, a dubious role of CII in FAO is suggested as a consequence (Figure 8a,b). Confused electron transfer pathways are described in Figure 8c (Supplement 2, Weblink #9) and Figure 8d (Supplement 3, Weblink #36). Lemmi et al (1990) noted: 'mitochondrial Complex II also participates in the oxidation of

fatty acids'. This holds for the oxidation of acetyl-Co in the TCA cycle, forming NADH and succinate with downstream electron flow through CI and CII, respectively, into the Q-junction (Figure 1). In contrast, electron transfer from FADH₂ formed during β-oxidation proceeds through electron transferring flavoprotein CETF and entry into the Q-junction independent of CII (Figure 7).

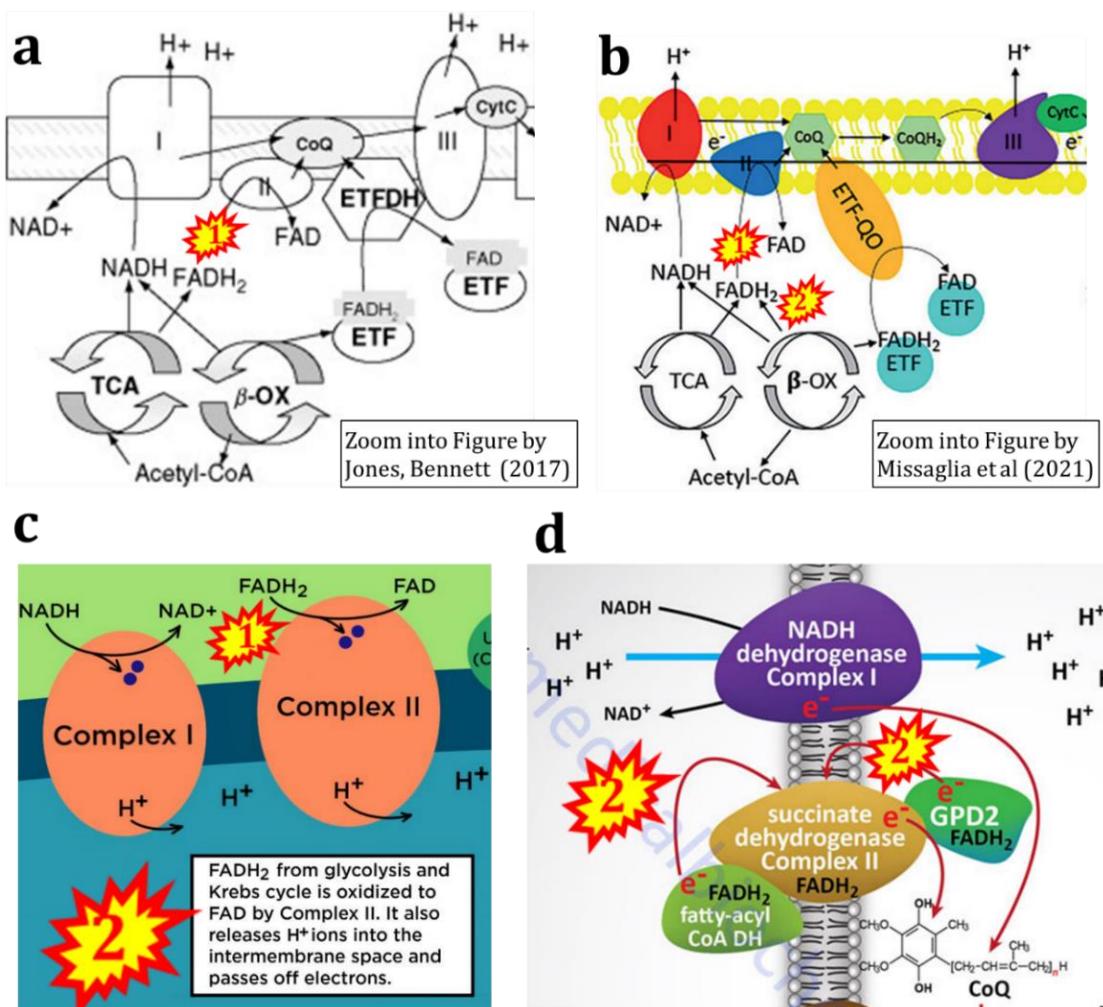


Figure 8. When FADH₂ is erroneously shown as a substrate of CII (1), a role of CII in oxidation of FADH₂ from glycolysis and fatty acid oxidation is suggested as a consequence (2). Zoom into figures by (a) Jones, Bennett (2017); (b) Missaglia et al (2021); (c) <https://www.expii.com/t/electron-transport-chain-summary-diagrams-10139> (accessed 2023-03-21); (d) <https://themedicalbiochemistrypage.org/oxidative-phosphorylation-related-mitochondrial-functions/> (accessed 2023-03-21).

4. Conclusions

The integration of FAO with the membrane-bound ETS (Wang et al 2019) has significant implications for understanding and treating disorders related to β-oxidation and oxidative phosphorylation. Clarification instead of perpetuation of Complex II ambiguities helps to maintain the high scientific standards required for translating knowledge on metabolism into clinical solutions for mitochondrial diseases.

Abbreviations

C1	Complex I	FAO	fatty acid oxidation
CII	Complex II	FMNH ₂	reduced flavin mononucleotide
CETF	electron transferring flavoprotein	NADH ₂	reduced nicotinamide adenine dinucleotide
FADH ₂	reduced flavin adenoside dinucleotide	TCA cycle	tricarboxylic acid cycle

Acknowledgements

I thank Luiza H Cardoso and Sabine Schmitt for stimulating discussions, and Paolo Cocco for expert help on the graphical abstract and Figure 1c. Contribution to the European Union's Horizon 2020 research and innovation program Grant 857394 (FAT4BRAIN).

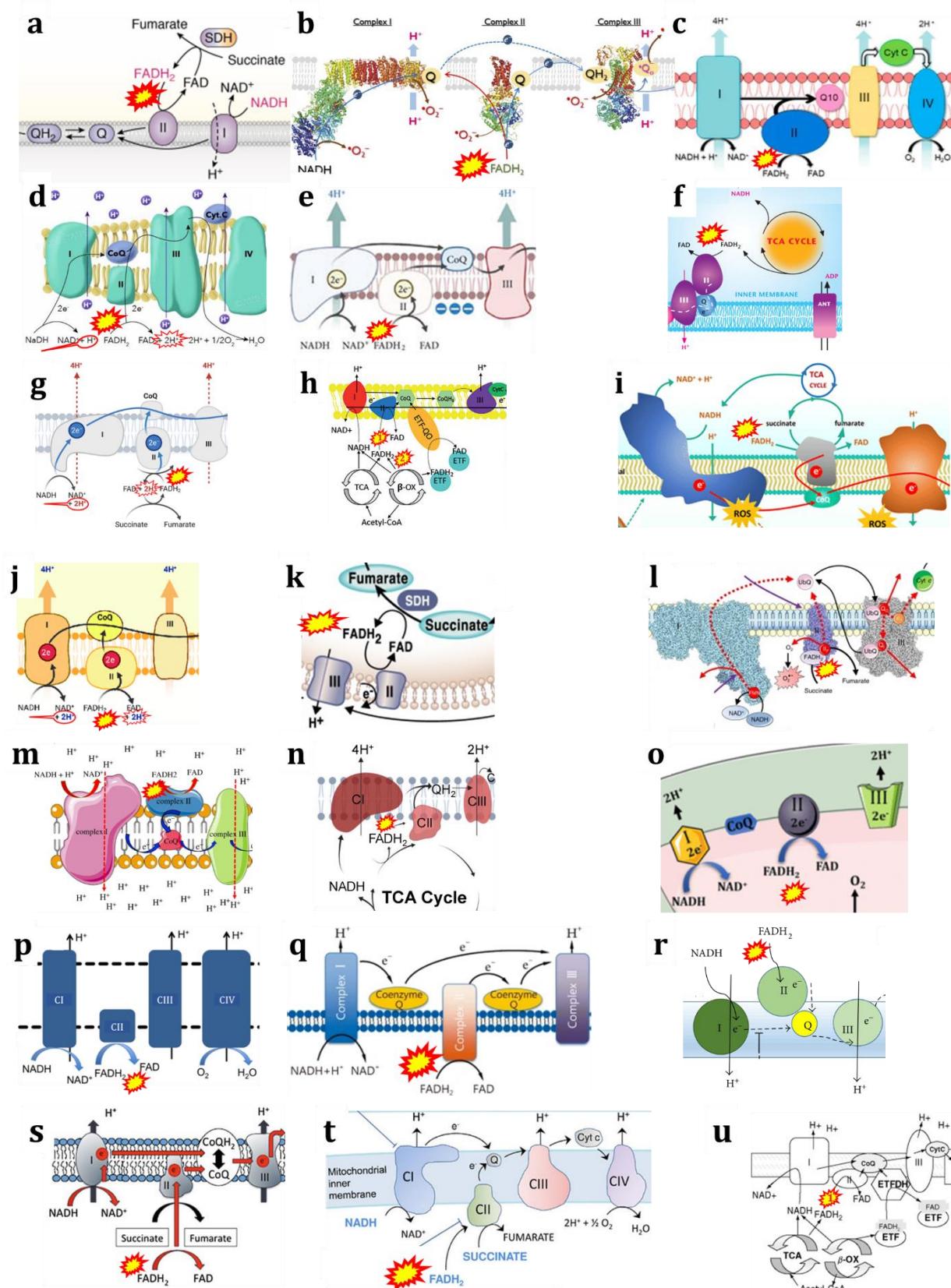
References

- Arnold PK, Finley LWS (2023) Regulation and function of the mammalian tricarboxylic acid cycle. <https://doi.org/10.1016/j.jbc.2022.102838>
- Brownlee M (2001) Biochemistry and molecular cell biology of diabetic complications. <https://doi.org/10.1038/414813a>
- Chandel NS (2021) Mitochondria. <https://doi.org/10.1101/cshperspect.a040543>
- Chen CL, Zhang L, Jin Z, Kasumov T, Chen YR (2022) Mitochondrial redox regulation and myocardial ischemia-reperfusion injury. <https://doi.org/10.1152/ajpcell.00131.2021>
- Cooper GM (2000) The cell: a molecular approach. 2nd edition. Sunderland (MA): Sinauer Associates Available from: <https://www.ncbi.nlm.nih.gov/books/NBK9885/>
- Chouchani ET, Pell VR, Gaude E, Aksentijević D, Sundier SY, Robb EL, Logan A, Nadtochiy SM, Ord ENJ, Smith AC, Eyassu F, Shirley R, Hu CH, Dare AJ, James AM, Rogatti S, Hartley RC, Eaton S, Costa ASH, Brookes PS, Davidson SM, Duchen MR, Saeb-Parsy K, Shattock MJ, Robinson AJ, Work LM, Frezza C, Krieg T, Murphy MP (2014) Ischaemic accumulation of succinate controls reperfusion injury through mitochondrial ROS. <https://doi.org/10.1038/nature13909>
- DeBerardinis RJ, Chandel NS (2016) Fundamentals of cancer metabolism. <https://doi.org/10.1126/sciadv.1600200>
- Fisher-Wellman KH, Neufer PD (2012) Linking mitochondrial bioenergetics to insulin resistance via redox biology. <https://doi.org/10.1016/j.tem.2011.12.008>
- Gnaiger E (1993) Efficiency and power strategies under hypoxia. Is low efficiency at high glycolytic ATP production a paradox? In: Surviving hypoxia: Mechanisms of control and adaptation. Hochachka PW, Lutz PL, Sick T, Rosenthal M, Van den Thillart G (eds) CRC Press, Boca Raton, Ann Arbor, London, Tokyo:77-109.
- Gnaiger E (2020) Mitochondrial pathways and respiratory control. An introduction to OXPHOS analysis. 5th ed. <https://doi.org/10.26124/bec:2020-0002>
- Gnaiger E et al - MitoEAGLE Task Group (2020) Mitochondrial physiology. <https://doi.org/10.26124/bec:2020-0001.v1>
- Grosholz ER (2007) Representation and productive ambiguity in mathematics and the sciences. Oxford Univ Press:312 pp.
- Hatefi Y, Haavik AG, Fowler LR, Griffiths DE (1962) Studies on the electron transfer system XLII. Reconstitution of the electron transfer system. [https://doi.org/10.1016/S0021-9258\(19\)73804-6](https://doi.org/10.1016/S0021-9258(19)73804-6)
- Hochachka PW, Somero GN (2002) Biochemical adaptation: mechanism and process in physiological evolution. Oxford Univ Press, New York:466 pp.
- Houten SM, Violante S, Ventura FV, Wanders RJ (2016) The biochemistry and physiology of mitochondrial fatty acid β-oxidation and its genetic disorders. <https://doi.org/10.1146/annurev-physiol-021115-105045>
- Jones PM, Bennett MJ (2017) Chapter 4 - Disorders of mitochondrial fatty acid β-oxidation.

- Elsevier In: Garg U, Smith LD , eds. Biomarkers in inborn errors of metabolism. Clinical aspects and laboratory determination:87-101. <https://doi.org/10.1016/C2014-0-03841-5>
- Lane N (2022) Transformer: the deep chemistry of life and death. Profile Books:400 pp. ISBN-10: 0393651487
- Lemmi CA, Pelikan PC, Geesaman B, Seamon E, Koyle M, Rajfer J (1990) Kinetics of cyclosporine A-induced inhibition of succinate-coenzyme Q dehydrogenase in rat renal cortical mitochondria. [https://doi.org/10.1016/0885-4505\(90\)90027-x](https://doi.org/10.1016/0885-4505(90)90027-x)
- Martínez-Reyes I, Chandel NS (2020) Mitochondrial TCA cycle metabolites control physiology and disease. <https://doi.org/10.1038/s41467-019-13668-3>
- Missaglia S, Tavian D, Angelini C (2021) ETF dehydrogenase advances in molecular genetics and impact on treatment. <https://doi.org/10.1080/10409238.2021.1908952>
- Murphy MP, O'Neill LAJ (2018) Krebs cycle reimagined: the emerging roles of succinate and itaconate as signal transducers. <https://doi.org/10.1016/j.cell.2018.07.030>
- Robb EL, Hall AR, Prime TA, Eaton S, Szibor M, Visconti C, James AM, Murphy MP (2018) Control of mitochondrial superoxide production by reverse electron transport at complex I. <https://doi.org/10.1074/jbc.RA118.003647>
- Schöpf B, Weissensteiner H, Schäfer G, Fazzini F, Charoentong P, Naschberger A, Rupp B, Fendt L, Bukur V, Giese I, Sorn P, Sant'Anna-Silva AC, Iglesias-Gonzalez J, Sahin U, Kronenberg F, Gnaiger E, Klocker H (2020) OXPHOS remodeling in high-grade prostate cancer involves mtDNA mutations and increased succinate oxidation. <https://doi.org/10.1038/s41467-020-15237-5>
- Tretter L, Patocs A, Chinopoulos C (2016) Succinate, an intermediate in metabolism, signal transduction, ROS, hypoxia, and tumorigenesis. <https://doi.org/10.1016/j.bbabi.2016.03.012>
- Tzagoloff A (1982) Mitochondria. Plenum Press, New York 342 pp.
- Wang Y, Palmfeldt J, Gregersen N, Makhov AM, Conway JF, Wang M, McCalley SP, Basu S, Alharbi H, St Croix C, Calderon MJ, Watkins S, Vockley J (2019) Mitochondrial fatty acid oxidation and the electron transport chain comprise a multifunctional mitochondrial protein complex. <https://doi.org/10.1074/jbc.RA119.008680>
- Yépez VA, Kremer LS, Iuso A, Gusic M, Kopajtich R, Koňářková E, Nadel A, Wachutka L, Prokisch H, Gagneur J (2018) OCR-Stats: Robust estimation and statistical testing of mitochondrial respiration activities using Seahorse XF Analyzer. <https://doi.org/10.1371/journal.pone.0199938>

Copyright: © 2023 The authors. This is an Open Access preprint (not peer-reviewed) distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original authors and source are credited. © remains with the authors, who have granted MitoFit Preprints an Open Access publication license in perpetuity.

Supplement Figure S1



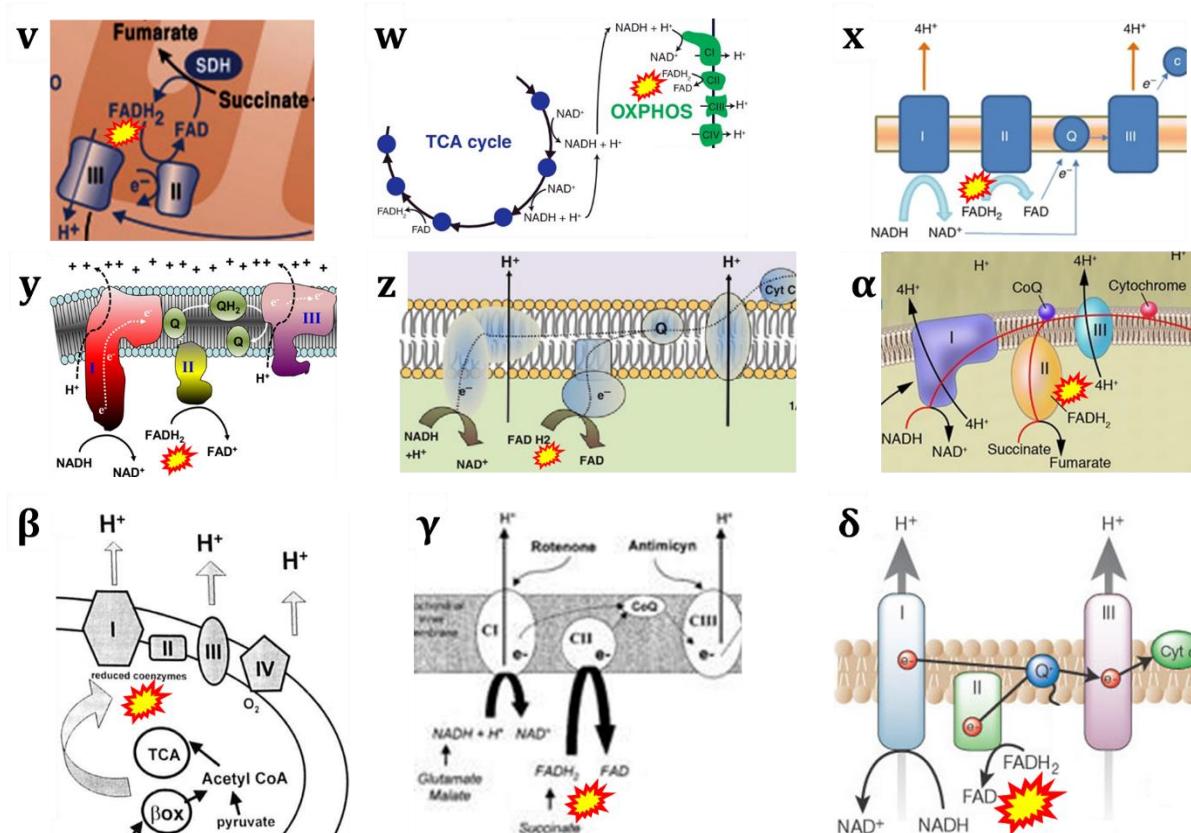


Figure S1. Complex II ambiguities in graphical representations on FADH₂ as a substrate of Complex II in the canonical forward electron transfer. Chronological sequence of publications from 2001 to 2023. See References for Figure S1.

References for Figure S1

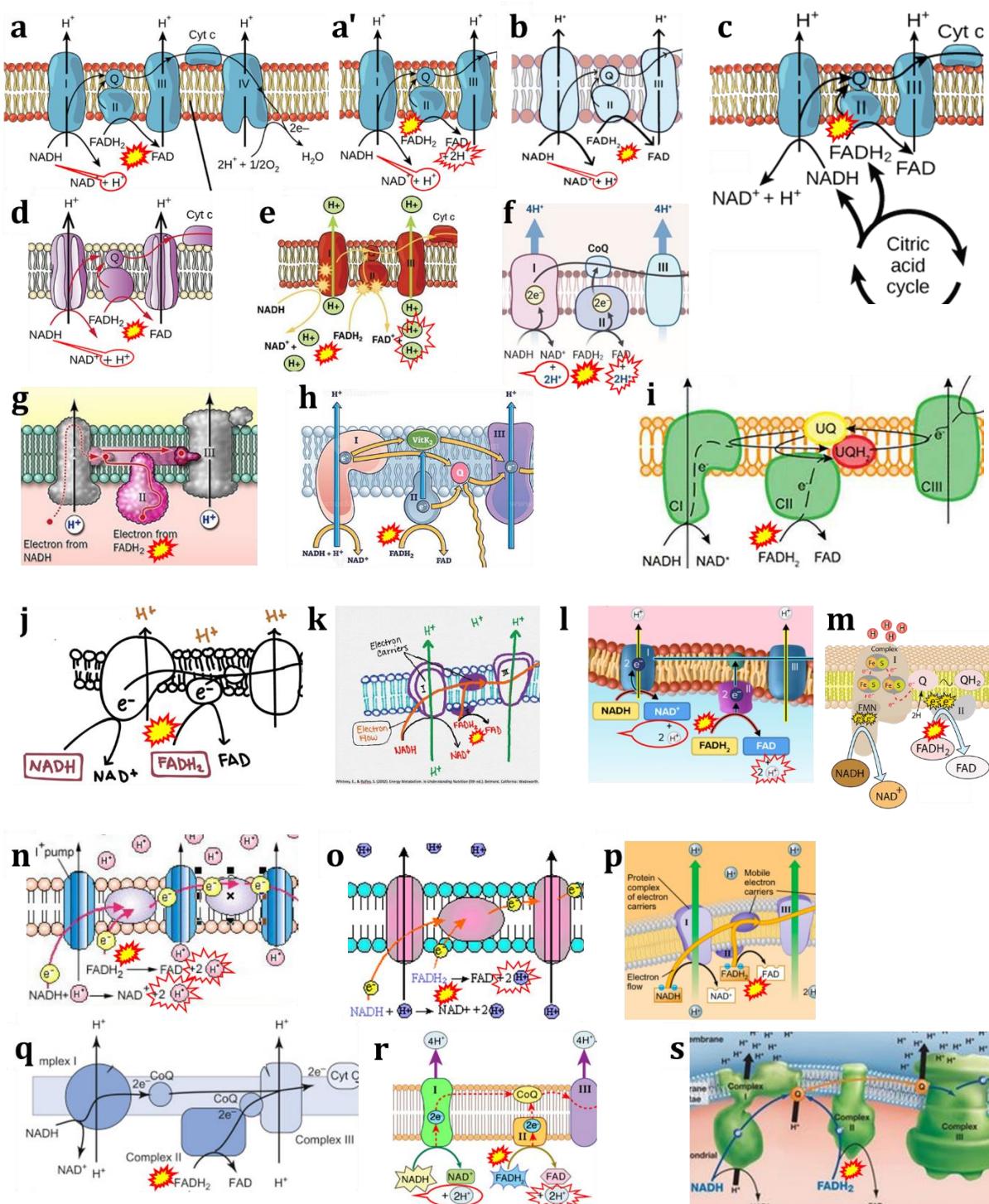
- a Arnold PK, Finley LWS (2023) Regulation and function of the mammalian tricarboxylic acid cycle. <https://doi.org/10.1016/j.jbc.2022.102838>
- b Chen CL, Zhang L, Jin Z, Kasumov T, Chen YR (2022) Mitochondrial redox regulation and myocardial ischemia-reperfusion injury. <https://doi.org/10.1152/ajpcell.00131.2021>
- c Turton N, Cufflin N, Dewsbury M, Fitzpatrick O, Islam R, Watler LL, McPartland C, Whitelaw S, Connor C, Morris C, Fang J, Gartland O, Holt L, Hargreaves IP (2022) The biochemical assessment of mitochondrial respiratory chain disorders. <https://doi.org/10.3390/ijms23137487>
- d Ahmad M, Wolberg A, Kahwaji CI (2022) Biochemistry, electron transport chain. StatPearls Publishing StatPearls [Internet]. Treasure Island (FL). <https://www.ncbi.nlm.nih.gov/books/NBK526105/>
- e Yuan Q, Zeng ZL, Yang S, Li A, Zu X, Liu J (2022) Mitochondrial stress in metabolic inflammation: modest benefits and full losses. <https://doi.org/10.1155/2022/8803404>
- f Chandel NS (2021) Mitochondria. <https://doi.org/10.1101/csphperspect.a040543>
- g Yin M, O'Neill LAJ (2021) The role of the electron transport chain in immunity. <https://doi.org/10.1093/fj.202101161R>
- h Missaglia S, Tavian D, Angelini C (2021) ETF dehydrogenase advances in molecular genetics and impact on treatment. <https://doi.org/10.1080/10409238.2021.1908952>
- i Gasmi A, Peana M, Arshad M, Butnariu M, Menzel A, Bjørklund G (2021) Krebs cycle: activators, inhibitors and their roles in the modulation of carcinogenesis. <https://doi.org/10.1007/s00204-021-02974-9>
- j Turton N, Bowers N, Khajeh S, Hargreaves IP, Heaton RA (2021) Coenzyme Q10 and the

- exclusive club of diseases that show a limited response to treatment. <https://doi.org/10.1080/21678707.2021.1932459>.
- k** Martínez-Reyes I, Chandel NS (2020) Mitochondrial TCA cycle metabolites control physiology and disease. <https://doi.org/10.1038/s41467-019-13668-3>
- l** Raimondi V, Ciccarese F, Ciminale V (2020) Oncogenic pathways and the electron transport chain: a dangeROS liaison. <https://doi.org/10.1038/s41416-019-0651-y>
- m** Morelli AM, Ravera S, Calzia D, Panfoli I (2019) An update of the chemiosmotic theory as suggested by possible proton currents inside the coupling membrane. <https://doi.org/10.1098/rsob.180221>
- n** Lewis MT, Kasper JD, Bazil JN, Frisbee JC, Wiseman RW (2019) Quantification of mitochondrial oxidative phosphorylation in metabolic disease: application to Type 2 diabetes. <https://doi.org/10.3390/ijms20215271>
- o** Sarmah D, Kaur H, Saraf J, Vats K, Pravalika K, Wanve M, Kalia K, Borah A, Kumar A, Wang X, Yavagal DR, Dave KR, Bhattacharya P (2019) Mitochondrial dysfunction in stroke: implications of stem cell therapy. <https://doi.org/10.1007/s12975-018-0642-y>
- p** Yépez VA, Kremer LS, Iuso A, Gusic M, Kopajtich R, Koňářková E, Nadel A, Wachutka L, Prokisch H, Gagneur J (2018) OCR-Stats: Robust estimation and statistical testing of mitochondrial respiration activities using Seahorse XF Analyzer. <https://doi.org/10.1371/journal.pone.0199938>
- q** Zhang H, Feng YW, Yao YM (2018) Potential therapy strategy: targeting mitochondrial dysfunction in sepsis. <https://doi.org/10.1186/s40779-018-0187-0>
- r** Roy Chowdhury S, Banerji V (2018) Targeting mitochondrial bioenergetics as a therapeutic strategy for chronic lymphocytic leukemia. <https://doi.org/10.1155/2018/2426712>
- s** de Villiers D, Potgieter M, Ambele MA, Adam L, Durandt C, Pepper MS (2018) The role of reactive oxygen species in adipogenic differentiation. https://doi.org/10.1007/5584_2017_119
- t** Polyzos AA, McMurray CT (2017) The chicken or the egg: mitochondrial dysfunction as a cause or consequence of toxicity in Huntington's disease. <https://doi.org/10.1016/j.mad.2016.09.003>
- u** Jones PM, Bennett MJ (2017) Chapter 4 - Disorders of mitochondrial fatty acid β -oxidation. Elsevier In: Garg U, Smith LD , eds. Biomarkers in inborn errors of metabolism. Clinical aspects and laboratory determination:87-101. <https://doi.org/10.1016/C2014-0-03841-5>
- v** DeBerardinis RJ, Chandel NS (2016) Fundamentals of cancer metabolism. <https://doi.org/10.1126/sciadv.1600200>
- w** Nsiah-Sefaa A, McKenzie M (2016) Combined defects in oxidative phosphorylation and fatty acid β -oxidation in mitochondrial disease. <https://doi.org/10.1042/BSR20150295>
- x** Prochaska LJ, Cvetkov TL (2013) Mitochondrial electron transport. In: Roberts, G.C.K. (eds) Encyclopedia of Biophysics. Springer, Berlin, Heidelberg. https://doi.org/10.1007/978-3-642-16712-6_25
- y** Fisher-Wellman KH, Neufer PD (2012) Linking mitochondrial bioenergetics to insulin resistance via redox biology. <https://doi.org/10.1016/j.tem.2011.12.008>
- z** Benard G, Bellance N, Jose C, Rossignol R (2011) Relationships between mitochondrial dynamics and bioenergetics. In: Lu Bingwei (ed) Mitochondrial dynamics and neurodegeneration. Springer ISBN 978-94-007-1290-4:47-68.
- α** Nussbaum RL (2005) Mining yeast in silico unearths a golden nugget for mitochondrial biology. <https://doi.org/10.1172/JCI26625>
- β** Sanchez H, Zoll J, Bigard X, Veksler V, Mettauer B, Lampert E, Lonsdorfer J, Ventura-Clapier R (2001) Effect of cyclosporin A and its vehicle on cardiac and skeletal muscle mitochondria: relationship to efficacy of the respiratory chain. <https://doi.org/10.1038/sj.bjp.0704129>
- γ** Himms-Hagen J, Harper ME (2001) Physiological role of UCP3 may be export of fatty acids from mitochondria when fatty acid oxidation predominates: an hypothesis. <https://doi.org/10.1177/153537020122600204>
- δ** Brownlee M (2001) Biochemistry and molecular cell biology of diabetic complications.

<https://doi.org/10.1038/414813a>

- Copied by: Arden GB, Ramsey DJ (2015) Diabetic retinopathy and a novel treatment based on the biophysics of rod photoreceptors and dark adaptation. Webvision In: Kolb H, Fernandez E, Nelson R, eds. Webvision: The organization of the retina and visual system [Internet]. Salt Lake City (UT): University of Utah Health Sciences Center; 1995-.
<https://www.ncbi.nlm.nih.gov/books/NBK310272/>

Supplement Figure S2



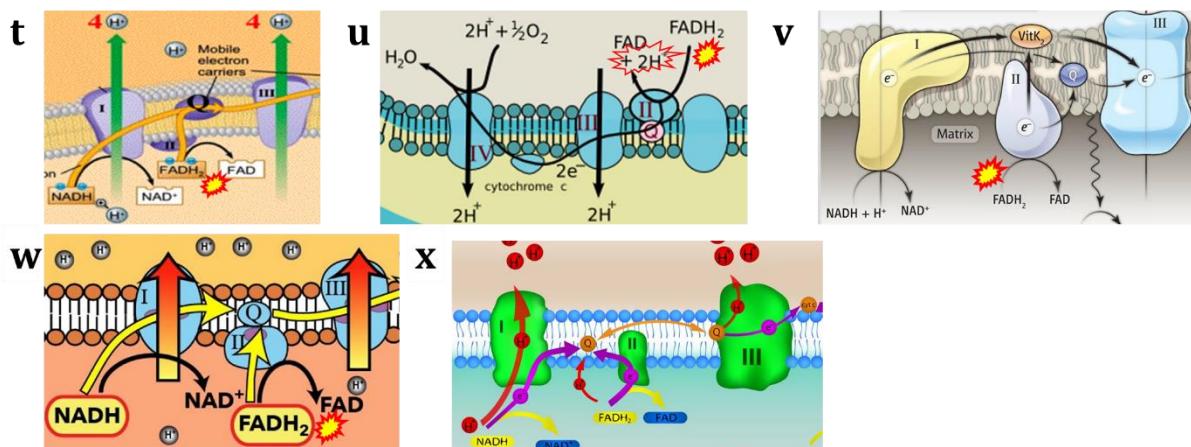


Figure S2. Complex II ambiguities in graphical representations on FADH₂ as a substrate of Complex II in the canonical forward electron transfer. Weblinks (#): a 1-5; a' 6-7; b 8; c 1, 6, 7, 9; d 10; e 4, 9, 11-16; f 17-18; g 19; h 20-21; i 22; j 6-7; k 9; l 23; m 24; n 25; o 26; p 27; q 28; r 29; s 30; t 31; u 9, 32; v 33; w 34; x 15, 17.

Weblinks for Figure S2 (retrieved 2023-03-21)

- 1 <https://openstax.org/books/biology/pages/7-4-oxidative-phosphorylation> - OpenStax Biology (CC BY 3.0) - Fig. 7.10 / Fig. 7.12
- 2 <https://opentextbc.ca/biology/chapter/4-3-citric-acid-cycle-and-oxidative-phosphorylation/> - Concepts of Biology - 1st Canadian Edition by Charles Molnar and Jane Gair - Fig. 4.19a
- 3 [https://bio.libretexts.org/Bookshelves/Introductory_and_General_Biology/Book%3A_General_Biology_\(Boundless\)/07%3A_Cellular_Respiration/7.11%3A_Oxidative_Phosphorylation_-_Electron_Transport_Chain](https://bio.libretexts.org/Bookshelves/Introductory_and_General_Biology/Book%3A_General_Biology_(Boundless)/07%3A_Cellular_Respiration/7.11%3A_Oxidative_Phosphorylation_-_Electron_Transport_Chain) - LibreTexts Biology - Figure 7.11.1
- 4 <https://courses.lumenlearning.com/wm-biology1/chapter/reading-electron-transport-chain/> - lumen Biology for Majors I - Fig. 1 / Fig. 3
- 5 <https://www.pharmaguideline.com/2022/01/electron-transport-chain.html> - Pharmaguideline
- 6 <https://www.khanacademy.org/science/ap-biology/cellular-energetics/cellular-respiration-ap/a/oxidative-phosphorylation-etc> - Khan Academy - Image modified from "Oxidative phosphorylation: Figure 1," by OpenStax College, Biology (CC BY 3.0) / Image modified from "Oxidative phosphorylation: Figure 3," by Openstax College, Biology (CC BY 3.0)
- 7 <https://learn.saylor.org/mod/page/view.php?id=32815> - Saylor Academy
- 8 <https://jackwestin.com/resources/mcat-content/oxidative-phosphorylation/electron-transfer-in-mitochondria> - Jack Westin MCAT Courses
- 9 <https://www.expii.com/t/electron-transport-chain-summary-diagrams-10139> - expii - Image source: By CNX OpenStax / By OpenStax College CC BY 3.0, via Wikimedia Commons / Whitney, Rolfe 2002 / By User:Rozzychan CC BY-SA 2.5, via Wikimedia Commons
- 10 <https://www.labxchange.org/library/items/lb:LabXchange:005ad47f-7556-3887-b4a6-66e74198fbef:html:1> - Labxchange - Figure 8.15 credit: modification of work by Klaus Hoffmeier
- 11 <https://commons.wikimedia.org/w/index.php?curid=30148497> - wikimedia 30148497 - Anatomy & Physiology, Connexions Web site. <http://cnx.org/content/col11496/1.6/>, 2013-06-19
- 12 <https://biologydictionary.net/electron-transport-chain-and-oxidative-phosphorylation/> - biologydictionary.net 2018-08-21
- 13 <https://www.quora.com/Why-does-FADH2-form-2-ATP> - Quora

[d&ved=2ahUKEwjilKSKpOX9AhWjmycCHbvGC34QMygWegUIARDWAQ](https://www.youtube.com/watch?v=d&ved=2ahUKEwjilKSKpOX9AhWjmycCHbvGC34QMygWegUIARDWAQ) - YouTube
sciencemusicvideos - Uploaded 2014-08-19

Supplement S3

Weblinks on FAO and CII (retrieved 2023-03-21)

- 35 <https://conductscience.com/electron-transport-chain/> - Conduct Science: "In Complex II, the enzyme succinate dehydrogenase in the inner mitochondrial membrane reduce FADH₂ to FAD⁺. Simultaneously, succinate, an intermediate in the Krebs cycle, is oxidized to fumarate." - Comments: FAD does not have a positive charge. FADH₂ is the reduced form, it is not reduced. And again: In CII, FAD is reduced to FADH₂.
- 36 <https://themedicalbiochemistrypage.org/oxidative-phosphorylation-related-mitochondrial-functions/> - The Medical Biochemistry Page: 'In addition to transferring electrons from the FADH₂ generated by SDH, complex II also accepts electrons from the FADH₂ generated during fatty acid oxidation via the fatty acyl-CoA dehydrogenases and from mitochondrial glycerol-3-phosphate dehydrogenase (GPD2) of the glycerol phosphate shuttle' (Figure 8d).
- 37 <https://www.chem.psu.edu/courses/chm333/Spring%202013/Lectures/Spring%202013%20Lecture%2037%20-%2038.pdf> - CHM333 LECTURES 37 & 38: 4/27 – 29/13 SPRING 2013 Professor Christine Hrycyna - Acyl-CoA dehydrogenase is listed under 'Electron transfer in Complex II'.