



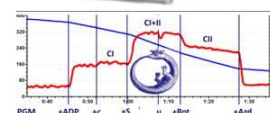
## O2k-Workshops

**IOC106.** Mitochondrial Physiology Network 20.10(01):1-8 (2015)  
Updates: [http://wiki.orooboros.at/index.php/MiPNet20.10\\_IOC106\\_Schroecken](http://wiki.orooboros.at/index.php/MiPNet20.10_IOC106_Schroecken)

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# 106<sup>th</sup> International Workshop on O2k high-resolution respirometry and O2k-Fluorometry

**2015 October 06-11**  
**Schröcken, Vorarlberg, Austria**



The **106<sup>th</sup> Workshop on O2k high-resolution respirometry O2k-Workshops HRR** (HRR) is the **34<sup>th</sup>** International Oxygraph Course held in Schroecken since 1988. An overview is provided of the **Oxygraph-2k, TIP2k, and O2k-Fluo LED2-Module**, with real-time analysis by **DatLab 6.2 (new)**. A demo experiment illustrates the principle and shows the unique advantage of simultaneous monitoring of oxygen concentration, respiration and **hydrogen peroxide production**. Cryopreserved cells are used as a biological reference sample (MitoFit proficiency test). Respiration and **mt-membrane potential** will be determined in hands-on experiments with cardiac tissue homogenate.

**Instrumental setup** and service of the polarographic oxygen sensor (**OroboPOS**) are demonstrated, followed by hands-on practice in 10 teams. In the evenings, general mitochondrial topics are covered; abstracts and experimental experiences are presented by participants.

IOC participants asked invariably for detailed discussions of protocol design. The **Blue Book** provides a basic introduction to mitochondrial physiology, complemented by overview presentations with examples, including **DatLab Analysis** of demo files. **Instrumental quality control** is a fundamental component of HRR and will be put to the practical test in teams using seven O2k (14 chambers). **O2k-MultiSensor** and particularly O2k-Fluorometry has become an integral part of the O2k-Workshop. Optimization of protocol design for various MultiSensor applications helps to critically evaluate basic principles of mitochondrial physiology. You will also see the **Titration-Injection microPump TIP2k** with feedback-control in action and practice its simple and automatic operation.

Lunch breaks provide an opportunity for relaxing walks and talks, enjoying the refreshing scenery of the secluded alpine environment, offer a visit to the Alpmuseum, or give sufficient spare time for individual practice.



## Lecturers and tutors

<a href="#">Doerrier Carolina</a>	Mitochondrial Application Specialist, NextGen-O2k, OROBOROS
<a href="#">Gnaiger Erich</a>	CEO, OROBOROS
<a href="#">Hoppel Florian</a>	PhD Student, MitoFit, OROBOROS
<a href="#">Laner Verena</a>	Chief Operating Officer (COO), OROBOROS
<a href="#">McManus Meagan</a>	O2k-Network Lab: <a href="#">US PA Philadelphia Wallace DC</a>
<a href="#">Sumbalova Zuzana</a>	O2k-Network Lab: <a href="#">SK Bratislava Sumbalova Z</a> ; OROBOROS

## Programme IOC106

### 1 Tuesday, Oct 06

\* printed in workshop materials

	Arrival	Weblink
15:00	<b>Arrival in Bregenz:</b> Meeting point Bregenz train station at 3:00 pm; approx. 1 hour bus drive to Schröcken and Hochtannberg (Salober). Transfer/walk to Hotel Körbersee.	<a href="#">IOC-travel</a>
18:30	<i>Welcome reception at Hotel Körbersee</i>	<a href="#">Schroecken</a>
19:00	<i>Dinner</i>	
20:30-21:00	<b>Get-together:</b> introduction of participants and their research interests – define working groups	<a href="#">IOC106</a>

### 2 Wednesday, Oct 07

	Workshop 1	Weblink
07:30-08:30	<i>Breakfast</i>	
08:30-09:30	<b>O2k instrumental setup</b> – overview with video clips	<a href="#">O2k-Manual</a>
	<b>Hands-on (10 groups)</b>	Special task group:
	<b><u>O2k instrumental setup</u></b>	<b><u>SUIT protocols</u></b>
09:30-10:15	Groups 1-5	<a href="#">O2k-Start</a>
10:15-11:00	Groups 6-10	<a href="#">POS Service</a>
11:00	<i>Coffee / Tea</i>	
11:30-12:00	<b>Get O2k-Connected with OROBOROS:</b> a guided tour to the O2k	<a href="#">Oxygraph-2k</a>
12:00	<i>Lunch packages/ walk &amp; talk / alternative: individual O2k-tasks</i>	
14:30-15:30	<b>Instrumental quality control 1:</b> O <sub>2</sub> calibration and the O2k quality control system	<a href="#">Gnaiger 2008 POS*</a> <a href="#">O2k-Calibration</a>
15:30	<i>Coffee / Tea</i>	
16:00-16:30	<b>Instrumental quality control 2:</b> Analysis of oxygen flux; O2k-background test with the TIP2k	<a href="#">O2-Flux Analysis</a>
16:30-18:00	<b>Hands-on (7 groups): O2k calibration and background test:</b> air saturation to zero oxygen concentration; or for permeabilized muscle fibres in the high-oxygen range of 500 - 200 µM. O2k-background with automatic TIP2k or manual titrations. Special interest group: O2k-background with TPP <sup>+</sup> electrodes	<a href="#">O2k-Background</a> <a href="#">TIP2k User Manual</a>
18:30	<i>Dinner</i>	
20:00-21:00	<b>DatLab analysis and group reports:</b> O2k-calibration and instrumental background	<a href="#">POS-Calibration-SOP</a> <a href="#">O2 Background</a>

### 3 Thursday, Oct 08

Workshop 2		Weblink
07:30-08:30	<i>Breakfast</i>	
08:30-09:15	<b>DatLab O<sub>2</sub> flux analysis:</b> Flux per volume, flux per mass, flow per cell, flux control ratio, flux control factor	<a href="#">Glossary:</a> <a href="#">Respiratory states</a>
09:15-10:00	<b>DatLab guide through the menus:</b> DL-Demo files and DL-Excel templates	<a href="#">DatLab Guide</a>
10:00	<i>Coffee / Tea</i>	
10:00-10:45	<b>Hands-on (7 groups) getting started with an O2k-experiment:</b> washing, stirrer test, air calibration	<a href="#">O2k-Calibration</a>
10:45-12:00	<b>O2k-Demo experiment:</b> Respiration of intact cells: Simultaneous measurement of oxygen consumption ( <a href="#">O2k-Core</a> ) and H <sub>2</sub> O <sub>2</sub> production ( <a href="#">O2k-Fluo LED2-Module</a> )	<a href="#">Makrecka-Kuka 2015 Biomolecules</a>
12:00	<i>Lunch packages/ walk &amp; talk / alternative: individual O2k-tasks</i>	<a href="#">The Blue Book p 56*</a>
14:00-16:00	<b>Hands-on (7 groups). O2k-experiment:</b> Respiration with permeabilized cells: SUIT protocols with 7 Power-O2k	
16:00	<i>Coffee / Tea</i>	
16:30-18:00	<b>SUIT protocol and DatLab analysis with Excel templates</b>	<a href="#">DatLab Flux Analysis</a>
18:30	<i>Dinner</i>	
20:00-21:00	<b>O2k perspectives:</b> 10+5 min presentations of abstracts 1-6	<a href="#">IOC106 Abstracts</a> <a href="#">MiPNet20.10</a>

### 4 Friday, Oct 09

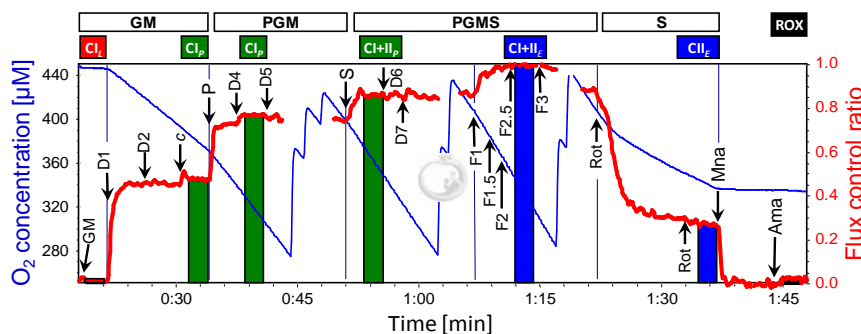
Workshop 3		Weblink
07:30-08:30	<i>Breakfast</i>	
08:30-09:30	<b>From isolated mitochondria to tissue fibres and tissue homogenate preparation:</b> The PBI-Shredder	<a href="#">MiPNet17.03 Shredder vs Fibres*</a>
09:30-10:00	<b>Hands-on (7 groups) getting started with an O2k experiment 2:</b> washing, stirrer test, air calibration	<a href="#">O2k-cleaning and ISS</a>
10:00	<i>Coffee / Tea</i>	
10:30-12:00	<b>O2k-experiment:</b> Respiration with tissue homogenate: Two SUIT protocols with 7 Power-O2k	<a href="#">Krumnschnabel 2013 Abstract MiP2013: 26-27*</a>
12:00	<i>Lunch packages/ walk &amp; talk / alternative: individual O2k-tasks</i>	
15:00-16:00	<b>DatLab analysis: hands-on in teams</b>	<a href="#">DatLab Flux Analysis</a>
16:00	<i>Coffee / Tea</i>	
16:30-17:15	<b>DatLab analysis: summary discussion</b>	<a href="#">DatLab Flux Analysis</a>
17:15-18:00	<b>O2k-Fluorometry demo: Calibration for H<sub>2</sub>O<sub>2</sub> production.</b>	<a href="#">MiPNet20.14 Ampl exRed H2O2-production</a>
18:30	<i>Dinner</i>	
20:00-21:00	<b>DatLab analysis: diagnosis of respiratory defects.</b>	

## 5 Saturday, Oct 10

Workshop 4		Weblink
07:30-08:30	Breakfast	
08:30-09:15	<b>Experimental design 1:</b> Substrate and coupling control of mitochondrial respiration - MitoPathways through CI&II	<a href="#">The Blue Book* pp 43-57</a>
09:15-10:00	<b>Experimental design 2:</b> Coupling control protocols: ROUTINE, LEAK, ETS, ROX	<a href="#">Cells: CCP Coupling control state</a>
10:00	Coffee / Tea	<a href="#">MiPNet18.10 O2kvsMultiwell*</a>
10:30-12:00	<b>O2k-MultiSensor overview and O2k-Fluorometry applications:</b> Amplex™ red, safranin and TMRM	<a href="#">MiPNet17.17 Ampl ex-Mouse-brain*</a>
12:00	Lunch packages	
12:30-15:30	Walk to the Alpmuseum: Guided tour and reception: 15 €	<a href="#">Alpmuseum*</a>
16:00	Coffee / Tea	
16:00-16:45	<b>Working groups:</b> Elaborate answers to the 'Questions for the O2k-Workshop'	<a href="#">IOC-Questions*</a>
16:45-17:30	<b>IOC-Questions - discussion of 'Answers' - technical support</b>	<a href="#">O2k-Technical support</a>
17:30-18:15	O2k-Network, MitoGlobal EAGLE and future O2k-plans (5+5 min) Anthony Molina, Meagan McManus, and participants to be nominated	<a href="#">O2k-Network</a>
18:15-18:45	<b>The O2k-Workshop continues with the Bioblast wiki - in the spirit of Gentle Science</b>	<a href="#">www.bioblast.at</a>
19:00	Dinner	
20:30-21:30	<b>Panel Discussion - Feedback</b> Farewell party	<a href="#">O2k-Feedback</a>

## 6 Sunday, Oct 12

Departure / Fish project	
Breakfast	
Early morning: Departure	



SUIT protocol with trout heart homogenate in a high oxygen concentration regime (MiR06Cr, 15 °C, reox with H<sub>2</sub>O<sub>2</sub>; [Krumchnabel 2013 Abstract MiP2013](#))\*.

## Participants

Participant	Institution
<a href="#">Ball Darran</a> **	<b>UK Southampton Grocott MP:</b> University of Southampton, Academic Unit of Cancer Sciences (UK)
<a href="#">Bezuidenhout Nicole</a> **	<b>ZA Cape Town Ojuka EO:</b> University of Cape Town, Institute of South Africa Newlands, ESSM UCT Dept of Human Biology Sports Science (ZA)
<a href="#">Coker Robert</a> *	<b>US AK Fairbanks Coker R:</b> University of Alaska, Institute for Arctic Biology (US)
<a href="#">Droescher Stephanie</a>	<b>AT Innsbruck OROBOROS:</b> OROBOROS INSTRUMENTS (AT)
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<a href="#">Garnham Jack</a> **	<b>UK Leeds Peers C:</b> LIGHT, University of Leeds, Cardiovascular & Neuronal Remodelling (UK)
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<a href="#">Haas Clarissa</a> ***	<b>BR Porto Alegre Souza DOG:</b> Universidade Federal do Rio Grande do Sul, Biochemistry Department (BR)
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<a href="#">Hansen Allan</a> **	<b>DK Copenhagen Poulsen HE:</b> Rigshospitalet Copenhagen, Clinical Pharmacology (DK)
<a href="#">Jackson Christopher</a>	<b>FI Helsinki Wartiovaara A:</b> Biomedicum Helsinki, Research Program for Molecular Neurology (FI)
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<a href="#">Pirinen Eija</a>	<b>FI Helsinki Pirinen E:</b> University of Helsinki, Research Program for Molecular Neurology (FI)
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<a href="#">Schmitt Sabine</a> *	<b>DE Munich Zischka H:</b> Helmholtz Zentrum München, Institute of Molecular Toxicology and Pharmacology (DE)
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<a href="#">Smith James</a> **	<b>ZA Cape Town Ojuka EO:</b> University of Cape Town, Institute of South Africa Newlands, ESSM UCT Dept of Human Biology Sports Science (ZA)
<a href="#">Takada Shingo</a> **	<b>JP Sapporo Yokota T:</b> Hokkaido University, Department of Cardiovascular Medicine, Hokkaido University Graduate School of Medicine (JP)
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<a href="#">West Malcolm</a> **	<b>UK Southampton Grocott MP:</b> University of Southampton, Academic Unit of Cancer Sciences (UK)

## MiPNet20.10 Abstracts IOC106: 10+5 min

### O2k perspectives

#### 1. Alteraas EK, Lindquist C, Berge RK, Bjorndal B (2015) OXPPOS capacity in mouse liver altered by modified fatty acids that inhibit or stimulate beta-oxidation. *Mitochondr Physiol Network 20.10.*

Non-alcoholic fatty liver disease (NAFLD) is the most common liver disease in developed countries, characterized with high accumulation of triglycerides in the liver. NAFLD is associated with obesity, insulin resistance and dyslipidemia, and has been linked to reduced mitochondrial function. Tetradecylthiopropionic acid (TTP) and Tetradecylthioacetic acid (TTA) are artificial fatty acids, shown to inhibit and increase mitochondrial  $\beta$ -oxidation respectively. They were used to investigate the effect of  $\beta$ -oxidation on mitochondrial respiration as well as important mitochondrial factors. In the present study, C57BL/6JBomTac male mice ( $n = 8-10$ ) were fed a low-fat diet for three weeks supplemented with TTP, TTA, or combined TTP and TTA. As expected,  $\beta$ -oxidation was significantly higher in liver homogenate from TTA-treated mice. TTP treatment decreased hepatic  $\beta$ -oxidation, although not significantly. In accordance with the tendency of inhibited  $\beta$ -oxidation, plasma carnitine and acetylcarnitine levels were lower in TTP-treated mice than in controls, while palmitoylcarnitine levels increased. Concomitantly, liver TAG and cholesterol levels were increased. In contrast, TTA increased plasma carnitine levels, but did not affect liver lipid levels. Interestingly, the combined intervention of TTP + TTA increased  $\beta$ -oxidation and prevented liver TAG and cholesterol accumulation, but despite this an accumulation of plasma palmitoylcarnitine and a decrease in acetylcarnitine was observed. Our results show significantly higher oxidative phosphorylation and maximal capacity of the electron transport system in liver samples from TTA-treated mice. This increase was mainly due to increased complex I activity. TTP did not significantly alter mitochondrial respiration. Strikingly, in mice treated with a combination of TTP and TTA, oxidative phosphorylation was significantly decreased. Our findings suggest that TTA impact the mitochondrial respiration and fatty acid catabolism positively, while TTP did not have any effect on the mitochondrial respiration. Combined, TTA was able to prevent the TTP-induced liver lipid accumulation, but this was linked to a reduced OXPPOS capacity, which could indicate mitochondrial malfunction. More analyses and studies should be done to elaborate this further.

#### 2. Power A, Crossman D, Hickey AJ, Ward M (2015) Investigating mitochondrial function in a rat model of right ventricular hypertrophy. *Mitochondr Physiol Network 20.10.*

Background: The heart is a highly aerobic organ with more than 90% of ATP regenerated by oxidative phosphorylation (OXPPOS) in the mitochondria. Mitochondrial dysfunction has been identified as a hallmark in the transition of compensatory hypertrophy to heart failure. However, the contribution of mitochondrial dysfunction to the contractile deficit is debated.

Objectives: To determine if mitochondrial energy supply compromises contractile function in right ventricular (RV) hypertrophy.

Methods: Rats were injected with monocrotaline (MCT) to induce pulmonary artery hypertension and RV hypertrophy, or saline (CON) for age-matched controls. Four weeks post-injection, multicellular cardiac trabeculae (length 2-3 mm, diameter 150-250  $\mu\text{m}$ ) were micro-dissected from the RV and mounted on an inverted microscope between a force transducer and a length changer. Steady-state force and intracellular  $\text{Ca}^{2+}$  transients were measured prior to saponin "skinning" of the sarcolemma to allow buffer access to the cytosol without damaging organelle membranes. Contraction and relaxation of the trabeculae was then assessed using buffered  $\text{Ca}^{2+}$  solutions, with and without exogenous ATP added to the superfusate.

Results: MCT trabeculae produced similar force to CON despite having lower  $\text{Ca}^{2+}$  transients. Following skinning, CON trabeculae showed no change in the maximum  $\text{Ca}^{2+}$ -activated force when exogenous ATP was removed from superfusate, while MCT trabeculae showed smaller contractures without exogenous ATP when stimulated with saturating  $\text{Ca}^{2+}$ .

Discussion: In this MCT model of compensated right ventricular hypertrophy there appears to be only a small contribution of mitochondrial dysfunction to contraction/relaxation when intracellular  $\text{Ca}^{2+}$  is controlled. This protocol can be used to further examine energy specific deficits in the failing heart, and to investigate the effects of drugs that modulate mitochondrial energy supply on contractile function.

**3. Scaini G, Gomes LM, Carvalho-Silva M, Arent CO, Mariot E, Quevedo J, Streck EL (2015) Co-administration of omega-3 fatty acids and mood stabilizers reverses the impairment of bioenergetic parameters induced by fenproporex administration in hippocampus of rats. Mitochondr Physiol Network 20.10.**

Bipolar disorder (BD) presents a complex alternating clinical course with recurrent mood changes including manic and depressive episodes. Moreover, studies show that changes in energy metabolism are involved in the pathophysiology of BD and that omega-3 ( $\omega$ 3) fatty acids have beneficial properties in the central nervous system, by modulate energy metabolism. Thus, in the present stud we evaluate the effect of  $\omega$ 3 fatty acids alone or in combination with lithium or valproate on bioenergetic parameters, namely respiratory complexes (CI, CII, CII-III, CIV), malate dehydrogenase, succinate dehydrogenase, citrate synthase and creatine kinase activities. We observed a significant decrease of succinate dehydrogenase, CII and CIV and creatine kinase activities in hippocampus of animals submitted to fenproporex administration, as compared to the control group. Additionally, the  $\omega$ 3 fatty acids in combination with VPA or Li were able to reverse the decrease in succinate dehydrogenase, CII and CIV activities. However, the decrease in CK activity was reversed only with  $\omega$ 3 fatty acids in association with VPA. The present findings support the idea that  $\omega$ 3 fatty acids plays an important role in the modulation of energy metabolism, and exercise essential antioxidant capacity in the central nervous system, suggesting that the  $\omega$ 3 fatty acids may be a possible contributing in BD therapy.

**4. Sharaf MS, Van den Heuvel MR, Stevens D, Kamunde C (2015) Mechanisms of mitochondrial import and interactions of zinc and calcium. Mitochondr Physiol Network 20.10.**

Zinc and calcium have highly interwoven functions that are essential for cellular homeostasis. Here, we studied the mechanisms of their import into mitochondria and interactions on oxidative phosphorylation (OXPHOS) and membrane potential ( $\Delta\Psi$ m). We showed that while the two cations permeated the mitochondrial inner membrane via different mechanisms they synergistically inhibited OXPHOS but antagonistically dissipated  $\Delta\Psi$ m. Ruthenium red completely prevented calcium-induced OXPHOS inhibition suggesting that exclusive entry of calcium into the matrix via mitochondrial calcium uniporter is a fundamental step for the functional impairment caused by this cation. Overall, these data indicate that interactions of zinc and calcium on mitochondrial function result from mechanisms other than competition for their uptake pathways.

**5. Volani C, Haschka D, Demetz E, Doerrier C, Gnaiger E, Weiss G (2015) Determination of mitochondrial respiration in peripheral blood mononuclear cells. Mitochondr Physiol Network 20.10.**

Mitochondria are dynamic organelles, involved in fundamental cell processes, including oxidative phosphorylation [1, 2]. Iron plays a decisive role in these processes because it is central part of mitochondrial enzyme complexes but also regulates citric acid cycle activity by modulating mitochondrial aconitase expression. Hence, imbalances of iron homeostasis impact on mitochondrial activity and, thus, on cell and organ functions [3]. So far, little information is available on how to best measure mitochondrial activity and its interaction with iron homeostasis in vivo; therefore we questioned whether determination of mitochondrial respiration in peripheral blood mononuclear cells (PBMCs) could be a good surrogate marker for that.

Human PBMCs were collected from buffy coats, purified cells ( $2 \times 10^6$  cells/ml) were resuspended in mitochondrial respiration medium (MiR05), and mitochondrial activity was assessed by high resolution respirometry (OROBOROS INSTRUMENTS, Austria). Moreover, to access the impact of iron on mitochondrial respiration we studied mitochondrial respiration in hearts and livers of mice, receiving either iron deficient- or standard iron-diet one week before being sacrificed. Organs were collected and stored in Custadiol prior to homogenization in MiR05. Mitochondrial routine respiration, complex I and II maximal oxidative phosphorylation together with non-coupled respiration of the homogenates were assessed at a final concentration between 1 and 2 mg.

Our ongoing experiments indicate that mitochondrial function testing can be successfully performed in human PBMCs as well as in mouse tissues. Analyses of organ samples from mice indicate that dietary iron supplementation leads to enhanced oxidative phosphorylation. Furthermore, it is plausible to hypothesize that PBMCs mitochondrial activity can reflect this organ increase. In conclusion, the use of high-resolution respirometry (OROBOROS INSTRUMENTS, Austria) represents a powerful and reliable tool to investigate mitochondrial respiration in PBMCs and tissues, and to systemically study the effects of iron homeostasis on mitochondrial function. Moreover, determination of mitochondrial function in PBMCs might provide useful information on mitochondrial activity in tissues.

## Accommodation and Location

**Hotel Körbersee** [www.koerbersee.at](http://www.koerbersee.at)  
T +43 5519 265; [hotel@koerbersee.at](mailto:hotel@koerbersee.at)



## More detail

Gnaiger E (2014) Mitochondrial pathways and respiratory control. An introduction to OXPHOS analysis. 4th ed. Mitochondr Physiol Network 19.12. OROBOROS MiPNet Publications, Innsbruck:80 pp. »[Bioblast link](#)«

**O2k-Manual** – <http://wiki.oroboros.at/index.php/O2k-Manual>

**O2k-Protocols** – <http://wiki.oroboros.at/index.php/O2k-Protocols>

**>1,500 O2k-Publications** – [www.bioblast.at/index.php/O2k-Publications](http://www.bioblast.at/index.php/O2k-Publications)

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[www.mitofit.org](http://www.mitofit.org)



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Cooperation and Feedback in Science



## NextGen-O2k

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O2k-Workshops are listed as [MitoGlobal Events](#)

